### **ROUTINE HEMATOLOGY** PROCEDURES

#### SPECIMEN COLLECTION

- Collection tubes are color coded
- Hematology uses the following
  - EDTA (ethylenediaminetetraacetic acid) chelates calcium - purple
  - Sodium Citrate binds calcium light blue
  - Lithium Heparin interacts with antithrombin III - green

#### PHLEBOTOMY COLLECTION

- EQUIPMENT Specimen Collection Tubes
- Needles
- Tourniquet
- Other gloves, alcohol, bandages, sharps containers



### VENIPUNCTURE



## CAPILLARY PUNCTURE



### MICROSCOPE

#### Resolution

Optical Defects



#### HEMACYTOMETER

#### Manual Counts



#### MANUAL WHITE CELL COUNTS

- 3% acetic Acid 1:100
- Total number of cells counted in 9 squares + 10% X 100 = cells/mm<sup>3</sup>
- 100 cells +10 X 100 = 11,000 cu mm or 11 X10<sup>9</sup>/L
- Note: using LeukoChek
- and neubaurer hemacytometer



### MANUAL RED CELL COUNTS

- Saline 0.85% 1:200
- Count 5 smaller squares within the center square
- Number of cells counted X dilution factor X 1/volumn = cells / mm<sup>3</sup> μL
- 100 cells x 200 x 1/0.04 = 500000 mm<sup>3</sup>
  500000mm<sup>3</sup> = 5 x10<sup>12</sup>/L



#### MANUAL PLATELET COUNTS

- 1% ammonium oxalate or acetic acid 1:100
- Count all areas in large center square
- Multiply the number of platelets counted times 1000
- Note: <u>Medix\* Thrombotic Pure Plus Test Kit</u> and neubauer hemacytometer

#### HEMOGLOBIN

- Cyanomethemoglobin
- 540 nm
- Lambert-Beer's Law
- Errors
  - □ †WBC Counts
  - Hgb S & C
  - Lipemia
  - Abnormal Globulins

### HEMATOCRIT

- Packed Cell Volume
  - Capillary tube
  - Centrifuge 10,000-15,000 g for 5 min
- Hgb x 3 = Hct



#### MEAN CELL VOLUME = MCV

- HCT (%)/ RBC (x 10<sup>12</sup>/L) X 10 = MCV
- 33/4.0 x 10 = 82.5 fl

#### MEAN CELL HEMOGLOBIN = MCH

Hgb (g/dl)/ RBC (x 10<sup>12</sup>/L) x 10 = MCH 11/4 x 10 = 27.5 pg

#### MEAN CELL HEMOGLOBIN CONCENTRATION = MCHC

Hgb(g/dl)/Hct(%) x 100 = MCHC

11/33 x 100 = 33.3 g/dl

#### SEDIMENTATION RATE

- Rate at which red cells settle from plasma
- Presence of inflammation
- Westergren or Wintrobe
- Things that effect sed rates
- Protein composition of plasma
- Size and shape of RBC
- RBC concentration



% reticulocytes = number of reticulocytes counted

#### RETICULOCYTE COUNT

- •
- Supravital Stain new methylene blue Miller Disk This disc consists of 2 squares as shown below in figure 5-2. The area of the smaller square (B) is a tenth that of square A. Therefore, if there are 40 red cells in square A, there should be four red cells present in square B. When employing this method to count reticulocytes, the red cells in square B are counted in successive fields on the slide, until a total of 500 red cells have been counted. At the same time, the reticulocytes in square A are enumerated. At the completion of the count, the count of the count, obtained in this way are divided by 50, in order to obtain the percent reticulocytes present in the blood. . Count retics in 5 fields of 200 RBC # or retic/ 10 = % retics





#### SOLUBILITY TEST FOR HEMOGLOBIN S

#### Sodium dithionite

NEGATIVE = Clear Solution POSITIVE = Turbid Solution





### NORMAL PERIPHERAL SMEAR

Microscopic evaluation of stained blood smears.











### METHODS

#### Manual

- Cover Glass
- Wedge Smear
- Automated
  Wedge Smear





#### DEMONSTRATION



## OPTIMAL BLOOD SMEAR CHARACTERISTICS

- Minimum Length 2.5 cm
- Gradual Transition from Thick to Thin
- Straight Feather Edge
- Margins Narrower than Slide
- No Streaks, Waves, or Troughs



### STAINING - ROMANOWSKY

- Methylene Blue
- Azure B (oxidation product of Methylene Blue)
- Eosin A or B
- Examples Wright, Wright-Giemsa, Leishman, May-Grunwald, Jenner



#### GOOD SMEAR

- Macroscopic Pinkish Purple in Color
- Microscopic
  - Cells Evenly Distributed
  - Areas between cells clearErythrocytes are orange red
  - Neutrophilic Granules are lilac
  - Eosinophilic Granules are red orange
  - Basophilic Granules are purplish black
  - Lymphocyte cytoplasm is blue
  - Leukocyte nucleoli are purple



#### CAUSES OF BAD SLIDES

- Excessively Blue or Dark
  - Prolonged Staining
  - Inadequate Washing
  - Too Alkaline Stain or Buffer
  - Thick Blood Smears
- Excessively Pink or Light
  - Insufficient Staining
  - Prolonged Washing
  - Too Acidic Stain or Buffer
- Presence of Precipitate
  - Unclean Slides
  - Drying during Staining Process
- Inadequate Filtration of Stain



### SCANNING THE SLIDE

# ABNORMALITIES AND EFFECT ON CELL COUNTS

- Smudge Cells- none
- Nucleated RBC increased WBC counts
- Platelet Clumps decreased platelet counts
- Platelet Satellitism decreased platelet counts
- RBC agglutination decreased RBC count
- Rouleaux none



#### SMUDGE CELL

Degenerating Leukocytes



### NUCLEATED RBC

Immature RBC



### PLATELET CLUMPS

 Clotted Specimen, EDTA induced clumping



#### PLATELET SATELLITISM

 Platelets surround leukocytes



#### ERYTHROCYTE AGGLUTINATION

RBC sticking together



#### ROULEAUX

 Stacking of RBC, looks like coins



#### ESTIMATING LEUKOCYTE COUNTS

# leukocytes counted in 5 fields(10X)/5 = A

A X 0.2  $\times 10^{9}$ /L = leukocytes x  $10^{9}$ /L

Example 200 leukocytes counted in 5 fields  $200/5 = 40 \times 0.2 = 8 \times 10^{9}/L$ 

#### ESTIMATING PLATELET COUNTS

# Platelets counted in 5 fields(100X)/5 = A

A X 15 x10<sup>9</sup>/L = platelets x 10<sup>9</sup>/L

Example 150 platelets counted in 5 fields  $150/5 = 30 \times 15 = 450 \times 10^{9}/L$ 

#### LEUKOCYTE COUNT CORRECTION FOR NUCLEATED RBC

Leukocyte count X 100/ 100 + # NRBC = Corrected Count

Example Leukocyte Count =  $15 \times 10^{9}$ /L Nucleated RBC per100 cells counted = 10

15 x 10<sup>9</sup>/L X 100 / 100 + 10 = 13.6 x 10<sup>9</sup>/L

#### AUTOMATED INSTRUMENTS

- Impedance Instruments
- Light Scatter Instruments



#### Review

- Blood Collection
- Manual counts for WBC, RBC, Platelets, HCT
- Understanding MCV, MCH, MCHC
- Other testing: sed rates, retic counts, solubility tests
- Blood Smears: Making, staining, characteristics, counting, abnormalities
- Instruments